

B2
CMBL
machinery (e.g., cysteine bridges, glycosylation and acylation patterns), etc.--

Please replace the paragraph on page 49, line 27 to page 50, line 2 with:

B3
--The *amdS* gene flanked by the 230 bp repeated sequences was obtained from pJRoy47 as a *SwaI/PmeI* fragment and inserted into *EcoRV/StuI* digested pDM156.2 to create pDM222.A (Figure 3). pDM222.A was digested with *EcoRI* and the 4.4 kb *EcoRI* fragment containing the *pyrG* deletion cassette was gel purified using QIAQUICK Gel Extraction Kit (Qiagen, Chatsworth, CA) prior to transformation.--

Please replace the paragraph on page 50, lines 3-12, with:

B4
--Spores of *Fusarium venenatum* MLY3 were generated by inoculating a flask containing 500 ml of RA sporulation medium with three 1cm² mycelia plugs from a minimal medium plate and incubating at 28°C, 150 rpm for 2 to 3 days. Spores were harvested through MIRACLOTH (Calbiochem, San Diego, CA) and centrifuged 20 minutes at 7000 rpm in a SORVALL RC-5B centrifuge (E. I. DuPont De Nemours and Co., Wilmington, DE). Pelleted spores were washed twice with sterile distilled water, resuspended in a small volume of water, and then counted using a hemocytometer.--

IN THE CLAIMS:

Please substitute the following amended claims for the pending claims having the same claim numbers:

1. (Amended.) A method of constructing and selecting or screening a library of polynucleotide sequences of interest in filamentous fungal cells, wherein the method comprises:

(a) transforming the fungal cells with a population of DNA vectors, wherein each vector comprises:

- B5
Sub
C1
- (i) a fungal selection marker polynucleotide sequence and a fungal replication initiating polynucleotide sequence, wherein the marker and the replication initiating sequence do not vary within the population; and
 - (ii) a polynucleotide sequence of interest, wherein there are vectors in the population that vary from other vectors in the population by carrying different versions of the polynucleotide sequence of interest;